


The gain of additional sampling methods for the fish-based assessment of large rivers

Petr Zajicek  <https://orcid.org/0000-0002-4523-384X>, Christian Wolter  <https://orcid.org/0000-0002-2819-2900>

DOI

[10.1016/j.fishres.2017.09.018](https://doi.org/10.1016/j.fishres.2017.09.018)

Original publication date

29 September 2017 (Available online)

Document version

Accepted manuscript

Published in

Fisheries Research

Citation (Vancouver)

Zajicek P, Wolter C. The gain of additional sampling methods for the fish-based assessment of large rivers. Fisheries Research. 2018;197:15-24.



1 The gain of additional sampling methods for the fish-based assessment of large rivers

2
3 Petr Zajicek¹ & Christian Wolter¹

4 ¹Leibniz-Institute of Freshwater Ecology and Inland Fisheries, Department Biology and Ecology of Fishes,
5 Mueggelseedamm 310, 12587 Berlin, Germany
6

7 Abstract

8 Fishes serve as indicators in ecological assessments of European large rivers. Electrofishing is
9 the standard fishing method although it is restricted to the shallow littoral shoreline. Fish
10 occurring in the open water zone of the main channel remain consequently underestimated.
11 Additional sampling methods that cover the mid-channel of rivers could close the
12 electrofishing-gap, but strengths, weaknesses and gains of both electrofishing and additional
13 sampling methods for fish-based assessments of large rivers have not been contrasted yet. We
14 analyzed a unique dataset consisting of 2,693 fish samplings in European large rivers and
15 compared electrofishing with the additional sampling methods trawling, seining, and drift-
16 netting. We compiled fish metrics commonly used in fish-based assessments yielded by the
17 different gears and highlight the differences in fish species, biodiversity metrics (Shannon
18 Index, Evenness, Simpson Index), the Fish Region Index (FRI) and densities of fish in
19 selected guilds (eurytopic, rheophilic, lithophilic, phytophilic, psammophilic, potamal) that
20 are considered indicative for the degradation of habitats in large rivers. Electrofishing yielded
21 overall highest numbers of species, biodiversity metrics and densities of fish guilds, except for
22 the number of migratory and Habitat Directive species, the FRI and densities of potamal fish.
23 The additional gears, predominantly trawling, captured additional rheophilic and lithophilic
24 species. Trawling also assessed most migratory and Habitat Directive species and yielded
25 higher densities of potamal fish as well as larger fish than electrofishing. Trawl catches
26 further estimated higher biodiversity compared to seining, while the latter yielded higher
27 densities of eurytopic, rheophilic, lithophilic and phytophilic fish. Drift-netting yielded the
28 lowest estimates overall but sample size was very low. We suggest that electrofishing is an
29 appropriate method to assess and evaluate the effects of hydromorphological degradation and
30 rehabilitation on fish, and to guide river management. It sufficiently well represents the
31 typical fish assemblage of large rivers despite its restriction to the shoreline. In contrast,
32 assessing specifically Habitat Directive, migratory and rare species, as well as obtaining
33 complete species inventories, e.g., for biodiversity assessments, requires complementary
34 sampling of the mid-channel of large rivers by additional gears such as trawling.
35

36 **Keywords:** Fishing gear comparison, trawl, seine, drift-net, electrofishing
37
38

39 Highlights:

- 40 ■ Shorelines and the mid-channel form two distinct meso-habitats in large rivers
 - 41 ■ Electrofishing is applied at the shoreline but well represents the fish assemblage
 - 42 ■ Electrofishing estimated higher biodiversity and density of habitat-sensitive fish
 - 43 ■ Additional gears that cover the mid-channel captured additional species
 - 44 ■ Trawling applied in the mid-channel captured higher densities of potamal fish
- 45

46 corresponding author:

47 Petr Zajicek

48 zajicek@igb-berlin.de

49 Leibniz-Institute of Freshwater Ecology and Inland Fisheries

50 Department Biology and Ecology of Fishes

51 Mueggelseedamm 310

52 12587 Berlin
53 Germany
54

55 **1. Introduction**

56 Representative sampling is a crucial challenge in ecological assessments of large rivers
57 (De Leeuw et al., 2007; Poikane et al., 2014), i.e., in rivers with a catchment size >10,000 km²
58 (Berg et al., 2004). Challenges arise from the pure size of the water body (Flotemersch et al.,
59 2011), the complexity of the riverine ecosystem (Ward et al., 2002) with its variety of habitat
60 structures (Loisl et al., 2013), the varying suitability and selectivity of different sampling
61 methods and the diversity of fish assemblages with broad requirements on specific habitats
62 (Penczak and Jakubowski, 1990). The shoreline and the open water zone of the main channel
63 are two distinct meso-habitats of large rivers. The littoral shoreline is rather shallow and
64 therefore has a great variety of differently structured micro-habitats such as sand banks,
65 gravel bars or areas loosely to densely colonized by emerged or submerged vegetation (Erős
66 et al., 2008; Lechner et al., 2014). Complex structures such as large wood provide refuge,
67 both for fish and prey organisms (Lynch and Johnson, 1989) and also aquatic vegetation and
68 can strongly influence fish community dynamics (Casselman and Lewis, 1996; Jacobsen and
69 Perrow, 1998; Weaver et al., 1997). Hence, highest fish production and diversity are observed
70 at the shoreline (Randall et al., 1996). The open water zone of the main channel is rather
71 unstructured with higher flow velocities, greater depths and it further covers the major part of
72 the river by both area and water volume (Szalóky et al., 2014). Though Wolter et al. (2004)
73 have shown that the open water zone of the main channel has distinct fish assemblages, its
74 importance as an relevant meso-habitat for riverine fishes (Loisl et al., 2013; Szalóky et al.,
75 2014), especially for potamal species (Wolter and Bischoff, 2001) has long been neglected
76 (Dettmers et al., 2001b; Galat and Zweimüller, 2001).

77 Electrofishing is a standard method to sample fish, even in large rivers (e.g., Beier et al.,
78 2007; Dußling, 2009; Aparicio et al., 2011). Electrofishing efficiency is however limited to
79 shallow areas (Bohlin et al., 1989) and decreases even in small streams with increasing river
80 width (Kennedy and Strange, 1981). It is well suited to sample complex habitat structures
81 such as aquatic vegetation or large wood, which harbor high concentrations of fish (Erős et
82 al., 2008; Lewin et al., 2014), but may be obstacles for most other sampling methods.
83 However, fish occurring in the open water zone of the main channel are underestimated by
84 electrofishing.

85 Additional methods such as trawling (e.g., Wolter et al., 2004), seining (e.g., Neebling and
86 Quist, 2011), gill-netting (e.g., Goffaux et al., 2005), drift-netting (e.g., Fladung, 2002), and
87 long-lining (e.g., Loisl et al., 2013) can be applied in the open water zone of the main channel
88 and could therefore be beneficial for the fish-based assessment of large rivers (Flotemersch et
89 al., 2011). However, besides long-lining, these fishing gears are prone to entanglements and
90 therefore less suitable for application in complex, structured habitats.

91 Biodiversity measures enhance understanding of the complex components driving
92 ecosystems (Morris et al., 2014). Biodiversity can however be biased because abundance of
93 species and densities of fishes can change in identical habitats during ontogeny (Blondel,
94 2003), between seasons (Dettmers et al., 2001a; Wolter and Bischoff, 2001) and even between
95 day and night (Erős et al., 2008; Wolter and Freyhof, 2004). Many fish species are further
96 either stationary or mobile throughout their lifecycle (Radinger and Wolter, 2014).
97 Composition of fish assemblages is accordingly variable even within identical habitats, which
98 makes assessments aiming to compare fish communities across large spatial extents rather
99 challenging.

100 Multiple sampling of identical sampling sites is beneficial (Dußling et al., 2004a; Kucera-
101 Hirzinger et al., 2008) to increase sample size and to minimize natural and temporal variation
102 due to, for example, sampling methodology, migration or habitat patterns (Wolter et al.,
103 2004). Repeated samplings over time (Magurran and Henderson, 2003) and over large spatial
104 extents (Tokeshi, 1993) further decrease sampling error and increase estimates of species
105 richness. On the other hand, repeated samplings lead to some challenges in statistical analyzes

106 (Poikane et al., 2014). Different approaches regarding sampling or analytical methodology
107 combined with variable fish traits can result in contrasting conclusions on ecological states
108 (Heino et al., 2013), requiring a certain standardization, especially when large-scale data are
109 considered.

110 The main objectives of this study were to evaluate commonly used fish sampling methods
111 and identify the gain of additional methods for the fish based assessment of large rivers while
112 accounting for the heterogeneity due to field sampling data. To achieve our objectives, we:

- 113 (i) compiled a dataset of 2,693 fish sampling occasions in European large rivers and
114 calculated various fish assemblage metrics commonly used in fish-based assessments;
- 115 (ii) compared fish metrics based on electrofishing with those based on trawling, seining,
116 and drift-netting in a first analysis comprising 849 fish samplings. Further, we tested
117 electrofishing against each additional method in three independent comparisons
118 standardized to similar sites sampled by both gears;
- 119 (iii) identified strengths, weaknesses and gains of applying additional sampling gears in
120 large rivers; and
- 121 (iv) evaluated whether electrofishing is sufficient for the fish-based assessment of large
122 rivers

123 We hypothesized that fish metrics depend on the sampling method used and that even
124 though additional sampling methods constitute valuable tools, the application of electrofishing
125 is superior for the fish-based assessment of large rivers. We further hypothesized that
126 additional sampling gears capture additional species and therefore complete the species
127 inventory, specifically concerning potamal fish. Thus, selection of sampling gears and use of
128 complementary sampling methods strongly depend on the study objectives. While obtaining
129 complete species inventories probably requires applying several sampling methods, the
130 evaluation of a rehabilitation structure in the littoral zone of a large river may not.

131 **2. Methods**

132 *2.1 The Large River Database (LRDB)*

133 The LRDB has been compiled within the EU project “Improvement and Spatial Extension
134 of the European Fish Index (EFI+, EC 044096) and further completed since. It consists of
135 2,693 sampling occasions from 358 sampling sites located in 16 European large rivers, i.e.,
136 rivers with a catchment size >10,000 km² (Berg et al., 2004). The LRDB is structurally
137 comparable to the Fish Database of European Streams, described in detail by Beier et al.
138 (2007). In contrast to the latter, it contains multiple samplings of identical sampling sites
139 using different gears, which allows for analysis of the improvement of fish metrics by
140 applying additional gears in large rivers.

141 The LRDB contains rivers Aller, Danube, Elbe, Ems, Havel, Ijssel, Lek, Meuse, Narew,
142 Oder, Rhine, Saale, Spree, Tisa, Vistula and Weser. River Danube and its tributary river Tisa
143 drain into the Black Sea. All other rivers drain into the North Sea or the Baltic Sea (Fig. 1).
144 Rivers were sampled in the main channel, in backwaters and in mixed locations (i.e., covering
145 both the straight channel and oxbows) across an average length of 2,221 m, 866 m and 951 m,
146 respectively. Assessments took place over several years (1996-2010), during different seasons
147 and a few samplings were conducted at night. The most frequent sampling methodology was
148 electrofishing (E: 1862) and trawling (T: 710), followed by seining (S: 48) and drift-netting
149 (D: 47). The remaining 26 samplings using gill-netting (23), long-lining (2) and fyke-netting
150 (1) had to be excluded from further analyses due to a lack of comparability. Fished length and
151 fished width had been recorded for each sampling occasion for electrofishing, trawling and
152 drift-netting and fished area is given for seining which allowed determining species densities
153 assessed by each method. Further, total length of captured fish had been recorded for some
154 samplings and species, which allowed to considering size selectivity between electrofishing
155 and trawling for frequently captured species.

156 2.2 *Data standardization protocol*

157 To standardize data, we selected only sampling occasions:

- 158 A. located in rivers draining into the North Sea and Baltic Sea. Rivers draining into the
159 Black Sea were excluded because they contain too distinct and more species-rich fish
160 communities biasing the comparisons;
- 161 B. covered a fished length of at least 400 m for electrofishing, trawling and drift-netting
162 to ensure that at least 95% of the species inventory were captured (Wolter et al.,
163 2004). Seining covered an area of at least 4000 m²;
- 164 C. captured at least 100 fish to fulfill national sampling standards (Dußling et al., 2004a)
165 while maintaining reasonable sample sizes for the gear comparisons;
- 166 D. conducted during daytime; and
- 167 E. conducted in the main channel.

168 The remaining dataset consisted of 849 samplings at 159 sites in 14 rivers. Electrofishing
169 (59.7%) and trawling (35.5%) were the most commonly applied gears followed by seining
170 (4.5%) and drift-netting (0.2%). This dataset was used for a preliminary pairwise comparison
171 between all gears. Further, three independent standardized datasets were created to compare
172 electrofishing with each additional gear:

- 173 1. trawling (ET; samplings: 446; sites: 17; rivers: 5; assessed 1997-2008);
174 2. seining (ES; samplings: 78; sites: 4; rivers: 1; assessed 1997-2004); and
175 3. drift-netting (ED; samplings: 10; sites: 1; rivers: 1; assessed 1997-2000).

176 The key condition for each of these three datasets was, in addition to standardization steps
177 A – E, that both methods compared were applied at least once at each sampling site. At the
178 single locations this ensures that the same fish assemblage was sampled and that observed
179 differences between gears might be attributed to method. Fig. 1 shows the locations of all
180 sampling sites. However, each of these three final datasets still consisted of inhomogeneous
181 sample sizes and contains confounding effects due to pseudo-replication, violating the
182 assumption of independence (i.e., clustered and nested data as well as repeated measurements;
183 Zuur and Ieno, 2016), which had to be accounted for in the statistical analyzes. These were
184 repeated samplings at same sampling sites, in different rivers (ET comparison only), during
185 different seasons and in different years.

186 2.3 *Data analyzes*

187 Gear contribution to the sampling results was assessed using fish assemblage metrics
188 commonly applied in fish-based assessments of rivers referring to species, biodiversity and
189 selected ecological guilds (Noble et al., 2007). All catches were standardized according to
190 length / area sampled as individuals per 100 m² for each sampling occasion prior to data
191 analysis. The standardized fish densities were used to calculate densities of ecological guilds
192 and the Fish Region Index of the whole sample according to Dußling et al. (2004b).

193 In addition to the total number of fish species (including lamprey species) captured in all
194 sampling occasions (= species inventory), we highlight the number of species that were
195 captured exclusively by the different gears. We further analyzed numbers of species and
196 proportions of fish in the total catches (PROP) that are migratory, protected or Habitat
197 Directive species (Council Directive 92/43/EEC, 1994), referred to as 'HD species'. The very
198 few reported hybrids between species were excluded from all analyses.

199 Species richness S as basic measure of biodiversity (Spellerberg and Fedor, 2003) was
200 determined for each sampling occasion. Further common biodiversity measures calculated
201 here were the Shannon Index and Evenness (Spellerberg, 2008) and the Simpson Index
202 (Somerfield et al., 2008). Each index was calculated for each sampling as follows:

203 Species richness S

$$S = \text{number of species}$$

204 Shannon Index H

$$H = - \sum \left(\frac{n_i}{N} \right) \log \left(\frac{n_i}{N} \right)$$

205 Evenness e

$$e = \frac{H}{\log S}$$

206 Simpson diversity Index D

$$D = 1 - \sum \left(\frac{n_i}{N} \right)^2$$

207 where n_i = number of individuals of a species i ; N = number of all individuals of all species.

208 We further analyzed the whole sample Fish Region Index (FRI_{total}), referred to as FRI
 209 further on, which is a fish-specific index for differences between river and stream regions
 210 (Dußling et al., 2004b). It characterizes fish species by means of their probabilities of
 211 occurrence in different river regions (Wolter et al., 2013) within the longitudinal river
 212 zonation (Illies, 1961) and takes values from three to eight (Dußling, 2009). For instance, a
 213 FRI of 7.00 corresponds to typical fish species of the metapotamal river region, respectively
 214 the common bream region (Dußling et al., 2004b). The FRI_{total} relates to the entire fish
 215 assemblage at a site and is particularly valuable for the assessment of large rivers because it
 216 rather sensitively indicates hydromorphological impacts related to river regulation,
 217 impoundments, but also rhithralisation effects (Wolter et al., 2013). The FRI_{total} was
 218 determined for each sampling occasion as:

219 Fish Region Index $FRI_{(total)}$

$$FRI_{(total)} = \frac{\sum_{i=1}^s \left(FRI_i \frac{n_i}{S^2 FRI_i} \right)}{\sum_{i=1}^s \frac{n_i}{S^2 FRI_i}}$$

220 where n_i = number of individuals of species i ; FRI_i = FRI of species i ; $S^2 FRI$ = variance of the
 221 FRI of species i . FRI_i and $S^2 FRI$ were retrieved from the literature (given below).

222 We selected the eurytopic and rheophilic habitat guilds as well as the lithophilic,
 223 psammophilic and phytophilic reproduction guilds and considered those as indicative guilds
 224 for environmental change (Welcomme et al., 2006) and hence valuable for assessments. The
 225 eurytopic guild represents generalist species and therefore mostly serves as indicator for
 226 degradation. In contrast, rheophilic species prefer running waters with higher flow patterns,
 227 i.e., benefit from natural flow dynamics. Rhithralisation can therefore also indicate
 228 degradation of the stagnant flow dynamics of the potamal regions of large rivers by decreased
 229 densities of eurytops and increased densities of rheophils. Lithophilic and psammophilic
 230 species essentially depend on spawning substrates that are maintained by hydromorphological
 231 processes and require coarse and fine substrate, respectively. Phytophilic species are obligate
 232 plant spawners depending on aquatic vegetation.

233 The assignment of fish species to guilds and to the species-specific FRI and $S^2 FRI$ (Table
 234 S1, supplementary information) primarily followed the classification provided by Scharf et al.
 235 (2011). We used Dußling et al. (2004b) and EFI+ Consortium (2009) for the remaining
 236 species. The calculation of FRI and $S^2 FRI$ of single species is provided in Wolter et al.
 237 (2013). We further analyzed the potamal guild as it represents species inhabiting primarily the
 238 open water zone of the main channel (Wolter and Bischoff, 2001). Species numbers and
 239 PROP were determined and densities of fish analyzed for each guild.

240 Within the standardized comparisons of ET and ES, we also analyzed fish densities of
 241 single species that were captured in at least 50% of all samplings with each gear (referred to
 242 as common species: *Abramis brama*, *Gymnocephalus cernuus*, *Leuciscus idus*, *Perca*
 243 *fluviatilis* and *Rutilus rutilus*). Within the ET comparison, we further analyzed size selectivity
 244 of electrofishing compared to trawling based on the total length of all measured fish of each
 245 common species. No length measurements of fish were available for the seine and drift-net
 246 catches.

247 2.4 Statistics

248 Mixed effects models were used for statistical analyses because they are robust to
249 inhomogeneous samples inherent in most field data and because they allow account to be
250 taken of random effects and unequal sample sizes (Zuur et al., 2009). Random effects
251 resemble potential confounding effects from stratified sampling in time or space that violate
252 the assumption of independence (Gonzales and Griffin, 2004). Random effects were site (ES
253 comparison), site nested in river (ET comparison) and season nested in year. Method was
254 treated as fixed factor in each model. Models' goodness of fit was assessed using the Akaike
255 Information Criterion (AIC, Akaike, 1981). Separate mixed effects models were fitted for
256 each ecological guild and biodiversity index. This resulted in 33 models, i.e., 11 preliminary
257 models comparing all gears amongst each other (ETSD, Table S7), 11 models for the
258 standardized ET comparison (Table S10) and 11 models for the standardized ES comparison
259 (Table S12). The standardized ED comparison was not considered for statistical analyzes due
260 to a small sample size (Table S13). P-values of ETSD models were adjusted using Tukey post
261 hoc tests (Tukey, 1949) for multiple comparisons (Table S8). For each model, marginal R^2
262 and conditional R^2 were calculated as the amount of explained variance by the fixed effect
263 (i.e., the method) and by the fixed and all random effects, respectively (Nakagawa and
264 Schielzeth, 2013). Additional models were applied as described above within the ET (5
265 models, Table S15) and ES (5 models, Table S17) comparisons to test for differences in
266 densities of common species. Differences in the total length of common species within the ET
267 comparison were tested accordingly (five models, Table S19), but also included the sampling
268 occasion as an additional random effect to account for sampling-based stratification of length
269 measurements.

270 Data were analyzed in R 3.3.1 (R Development Core Team, 2016). We used the function
271 *lmer* in the R package *lmerTest* (Kuznetsova et al., 2016), which depends on package *lme4*
272 (version 1.1-12; Bates et al., 2015) for fitting linear mixed models. Response variables were
273 log-transformed when non-normality or heteroscedasticity was observed in residual plots. All
274 response variables were modeled with a Gaussian error. Tukey post hoc tests were applied
275 using function *glht* in the R package *multcomp* (version 1.4-5; Hothorn et al., 2016). The
276 function *r.squaredGLMM* in the R package *MuMIn* (version 1.15.6; Barton, 2016) was used
277 to determine marginal and conditional R^2 . Statistical figures were plotted using the function
278 *lineplot.CI* in the R package *sciplot* (version 1.1-0; Morales et al., 2012). Fig. 1 was drawn
279 using ArcMap, version 10.2.2.

280 3. Results

281 3.1 Preliminary comparison of all gears

282 849 samplings at 159 sites in 14 large rivers yielded 503,593 fish of 66 species (including
283 three lamprey species, referred to as fish in the following; Table S2). Most common fish were
284 generalist species belonging to the eurytopic guild and represented >71% of the total catch.
285 Electrofishing estimated highest total numbers of all species. Additional gears estimated
286 higher PROP of eurytopic, phytophilic and potamal species and trawling captured one
287 migratory species more than electrofishing (Table 1).

288 Electrofishing estimated significantly higher (Table S8) species richness, Shannon Index,
289 Evenness, and Simpson Index and lowest FRI (Fig. 2) as well as significantly higher densities
290 of eurytopic, rheophilic, lithophilic and psammophilic fish (Fig. 3, Table S6). Density of
291 phytophilic fish was significantly higher for electrofishing compared to trawling. Trawling
292 and seining estimated significantly higher densities of potamal fish than electrofishing.
293 Trawling yielded significantly higher estimates of species richness, the Shannon Index,
294 Evenness, the Simpson Index compared to seining and drift-netting and further higher
295 densities of psammophilic fish compared to seining. Seining yielded significantly higher
296 densities of eurytopic, lithophilic and phytophilic fish compared to trawling.

297 3.2 Standardized gear comparisons

298 The ET comparison yielded 249,040 fish of 47 species (Table 1). All six species captured
299 exclusively with trawling were rheophilic and lithophilic (Table S3). Trawling captured more
300 rheophilic, lithophilic, migratory and HD species than electrofishing (Table 1). The ES
301 comparison yielded 39,389 fish of 33 species (Table 1). Seining captured two specimen of
302 *Salmo salar* that was not captured with electrofishing (Table S4). The ED comparison yielded
303 4,192 fish of 18 species (Table 1). Drift-netting captured one specimen of *Abramis ballerus*
304 that was not captured with electrofishing (Table S5). PROP of eurytopic, phytophilic and
305 potamal fish were higher for all additional gears compared to electrofishing (Table 1).

306 Electrofishing led to the highest total numbers of species, of species exclusively caught by
307 one method, the significantly highest species richness, Shannon Index, Evenness and Simpson
308 Index and lowest FRI (Fig. 4) as well as significantly highest densities of eurytopic,
309 rheophilic, lithophilic, phytophilic and psammophilic fish (Fig. 5) compared to trawling
310 (Table S10) and seining (Table S12). Identical trends were indicated compared to drift-netting
311 (Table S13). Trawling estimated significantly higher densities of potamal fish than
312 electrofishing.

313 Trawling and seining assessed significantly higher densities of the potamal species
314 *Abramis brama*, whereas densities of all remaining common species were significantly higher
315 for electrofishing (Fig. 6) compared to trawling (Table S15) and compared to seining (Table
316 S17). Total lengths of the common species *Abramis brama*, *Leuciscus idus*, *Perca fluviatilis*
317 and *Rutilus rutilus* were significantly higher when captured with trawling as compared to
318 electrofishing (Fig. 6, Table S19).

319 4. Discussion

320 Our study revealed that electrofishing captured most (94%) species across 849 samplings
321 and clearly outperformed the other gears by 30% (trawling), 48% (seining) and 80% (drift-
322 netting). Standardized comparisons validated that electrofishing captured more species than
323 any other gear as well as the highest number of species exclusively caught by a single method.
324 These findings clearly underline the well-known importance of the littoral zone for fish
325 (reviewed by Strayer and Findlay, 2010), combined with the superior efficiency of
326 electrofishing therein. Nevertheless, all fishing gears indicated typical fish assemblages of the
327 metapotamal river region that was characterized by generalist species and a FRI of around
328 seven (Dußling et al., 2004b).

329 The littoral zone along the shorelines provides integral resources for fish to reproduce
330 (diverse spawning substrates), hatch (reduced flow patterns), feed (diverse terrestrial and
331 aquatic food and prey items) and shelter (diverse physical structures). Most fish species are
332 therefore encountered at the littoral zone, at least during some parts of their life-cycle.
333 Biodiversity and fish density (Randall et al., 1996), also as a result of higher productivity
334 (Lewin et al., 2014), are therefore substantially higher in structured littoral habitats compared
335 to the structure-free open water zone. Therefore, the higher efficiency of electrofishing
336 compared to the additional gears demonstrated here does not only reflect differences in
337 selectivity between the compared gears, but rather differences between the meso-habitats
338 sampled by the gears. Thus, although electrofishing left a gap concerning the sampling of the
339 mid-channel, it well represented typical assemblages of large rivers by species numbers and
340 biodiversity and it also captured highest densities of fish guilds that are indicative for
341 hydromorphological degradation. As hydromorphological enhancements of the littoral zone
342 constitute key rehabilitation measures to restore degraded habitats for riverine fishes (Kail and
343 Wolter, 2011), electrofishing is likely more suitable to assess their success than other fishing
344 methods that are applied within the mid-channel.

345 Concomitantly to the shoreline, the mid-channel also constitutes a unique meso-habitat of
346 large rivers that provides a vast refuge for potamal species (Wolter and Bischoff, 2001).

347 Further, the mid-channel line typically provides higher flow velocities that constitute
348 important guiding currents for upstream migrating fish (Benitez et al., 2015) such as
349 anadromous salmonids (e.g., Kemp and O’hanley, 2010). The main currents in the mid-
350 channel are also utilized by drifting fish larva (Lechner et al., 2016; Zitek et al., 2004) as well
351 as downstream migrating species such as *Anguilla anguilla* when navigating to the sea (Piper
352 et al., 2015). Correspondingly, additional gears applied in the mid-channel estimated higher
353 PROP of potamal fish than electrofishing and also contributed additional migratory species to
354 the total species inventory. Additional gears are hence likely more suitable for the assessment
355 of management measures that target the restoration of longitudinal connectivity to promote
356 fish migration (e.g., Fullerton et al., 2010; Kemp and O’hanley, 2010).

357 All other gears captured additional species to electrofishing in standardized comparisons.
358 Species richness further showed that a high sampling effort is required with any gear to
359 capture the whole species inventory of large rivers (Dembkowski et al., 2012), because
360 species richness was relatively low for each sampling occasion compared to the total number
361 of species captured across all samplings with each method. Therefore, a combination of
362 sampling gears is highly beneficial to capture more species and to complete the species
363 inventory (Gutreuter et al., 1995; Clark et al., 2007; Eggleton et al., 2010). Assessments
364 aiming to determine the species inventory should accordingly apply various fishing gears
365 covering both the shoreline and the mid-channel of the main channel and also extent sampling
366 effort.

367 Trawling was the only fishing gear that estimated higher densities of the potamal guild
368 and that captured most additional species to electrofishing in standardized gear comparisons.
369 It seems therefore more suited than seining or drift-netting to be applied in addition to
370 electrofishing to assess the entire species inventory, the density of potamal fish and to
371 specifically capture rare and migratory species. Higher PROP and densities of potamal fish in
372 trawl catches further underline that potamal fish preferably move within the mid-channel
373 during daytime and are therefore less represented in daytime-electrofishing catches. Trawling
374 further captured larger fish of common species (except the small-growing *Gymnocephalus*
375 *cernuus*) than electrofishing. Both the meso-habitat and the gear-based selectivity of
376 electrofishing and trawling (e.g., Wolter and Freyhof, 2004) contribute to predominantly
377 larger fish captured by trawling because larger fish rather utilize the mid-channel section of
378 the main channel (Wolter and Bischoff, 2001) and to predominantly smaller fish captured by
379 electrofishing. Electrofishing however assessed higher densities of all common species,
380 except the potamal *Abramis brama*. Consequently, trawling estimates lower densities of larger
381 fish whereas electrofishing rather estimates higher densities of smaller fish. Trawling would
382 further also capture older fish of large-growing species whereas electrofishing would
383 underestimate the abundance of large fish in general and of older fish of large-growing
384 species. Both the meso-habitat and gear-based size-selectivity have further implications for
385 the assessment of biomass as rather many fish captured with electrofishing would have a
386 lower biomass than rather few fish captured with trawling. Further benefits of additional
387 methods such as trawling applied in combination with electrofishing are accordingly
388 complementary size and age spectra (Goffaux et al., 2005; Porreca et al., 2013; Wiley and
389 Tsai, 1983) as well as biomass estimates of fishes and fish assemblages.

390 Seining partly covered both the littoral and open water zone of the main channel, which
391 was well reflected in the fish metrics estimated. However, in Iowa’s (USA) nonwadeable
392 rivers Neebling and Quist (2011) assessed sampling effort and resulting species numbers
393 estimated with electrofishing, trawling and seining and concluded that seining was ineffective.
394 Seining was found to underestimate species numbers, abundances and catch per unit effort in
395 small streams (Poos et al., 2007; Wiley and Tsai, 1983) and to capture lower numbers of rare
396 species than electrofishing in a small river (Poesch, 2014). Our findings support the lower
397 suitability of seining to assess the species inventory of large rivers. Biodiversity estimates

398 obtained by seining were lower compared to both electrofishing and trawling. However,
399 seining may be valuable for assessing densities of eurytopic, rheophilic, lithophilic and
400 phytophilic fish within the littoral zone, especially in the absence of complex habitat
401 structures.

402 Drift-netting yielded consistently the lowest estimates of each fish metric assessed. These
403 findings might be not representative at all, because only two drift-netting samples from the
404 same day could be used in our analyses. However, 94% of the 47 drift-net samplings in our
405 database had to be excluded from the analyses because they captured less than 100 fish
406 (median area sampled 85.000 m²). This indicates that drift-netting captures rather low
407 numbers of fish. Nevertheless, drift-netting captured one additional migratory species
408 compared to electrofishing though its rare application in the standardized comparison which
409 shows that drift-netting can also have gains for the assessment of biodiversity and migratory
410 species. Apart from the low catch rates, the application of drift-netting is also restricted due to
411 typical uses of the river channel such as inland navigation. Most large rivers serve as
412 navigable waterways and intense ship traffic prevents the application of a floating net within
413 the fairway.

414 Densities, biodiversity and fish size were shown to largely depend on the meso-habitat
415 sampled and the sampling method applied therein. Therefore, researchers and managers
416 should carefully select meso-habitats and sampling gears according to the research objectives
417 (De Leeuw et al., 2007; Flotemersch et al., 2011) and explicitly refer to the meso-habitat
418 sampled as well as account for the benefits and limitations of the sampling gears used. In case
419 of applying complementary sampling gears in both meso-habitats, each meso-habitat should
420 be addressed separately to e.g., describe density, size and biomass of fish within the mid-
421 channel and at the shorelines while number of captured species can be pooled to characterize
422 the whole species inventory of large rivers.

423 Differences in selectivity caused by physico-chemical parameters between the compared
424 gears were not explicitly tested in this study (but accounted for in statistical analyzes by
425 including random effects) as fishing gears were not applied under experimental conditions
426 and as fishing gears were applied in different meso-habitats. Poos et al. (2007) did however
427 not find any indications for turbidity, dissolved oxygen and conductivity to account for
428 selectivity differences between electrofishing and seining in a small river. Nevertheless, each
429 fishing gear has potential selectivity restrictions associated with environmental conditions
430 during sampling. For instance, Lyon et al. (2014) reported that efficiency of electrofishing
431 decreased with turbidity caused by higher river discharge. Further, the application of trawling
432 is restricted within dry years if water levels are too low. Seine nets on the other hand are
433 difficult to handle if velocities are too high, generally restricting their application to low flow
434 conditions. Environmental variation can be minimized by selecting identical seasons and time
435 of the day for the sampling and further by repeating the sampling multiple times within a
436 season. From the analytical perspective, statistical methods such as mixed effects models
437 (Zuur et al., 2009) that allow to account for stratification of the samples (e.g., per year,
438 season, river, site or sample) help to reduce the accompanying uncertainties stemming from
439 e.g., varying environmental conditions that are inherent in field samplings covering large
440 spatio-temporal scales.

441 *4.2 Management recommendations*

442 The availability of two distinct meso-habitats in large rivers has far reaching implications
443 for the assessment of large rivers. Appropriate sampling strategies largely depend on the
444 research questions (De Leeuw et al., 2007; Flotemersch et al., 2011) and should follow
445 clearly-defined objectives as they constitute an integral part for the evaluation of river
446 restoration (Morandi et al., 2014). Gears that can sample complex structures and that are
447 applied at the shoreline of large rivers such as electrofishing are consequently more likely to

448 capture more fish and more species but smaller fish. Electrofishing is therefore well suitable
449 to reflect the typical fish assemblage of large rivers and performs superior to additional
450 methods in evaluating the success of hydromorphological restoration projects along the banks.
451 Complementary sampling gears applied in the mid-channel section are more likely to capture
452 fish and species that specifically utilize currents for navigation and dispersal as well as larger
453 fish. Additional gears may perform better than electrofishing in assessing the success of
454 projects aiming for the reestablishment of large migratory species, the restoration of
455 longitudinal connectivity or the facilitation of fish migration and dispersal. Any combination
456 of sampling gears covering both the shoreline and the main channel will perform superior
457 over single fishing methods (Gutreuter et al., 1995; Clark et al., 2007; Eggleton et al., 2010)
458 when assessments aim for a complete inventory of all species present at a site (biodiversity) or
459 for recording rare, endangered and migratory species (Lintermans, 2016) as well as to obtain
460 complementary size, biomass and age spectra. Trawling appeared as a more beneficial
461 addition to electrofishing than seining and drift-netting to capture specifically migratory and
462 rare species and potamal fish and hence to estimate biodiversity. However, each method
463 requires considerable sampling efforts to capture a substantial proportion of the species
464 inventory (Neebling and Quist, 2011). To facilitate large scale assessments, sampling gears
465 need to be applied consistently (Goffaux et al., 2005) within similar meso-habitats and under
466 comparable environmental conditions.

467 **Acknowledgements**

468 This study and its extensive data compilation received support from the EU FP6 Project EFI+
469 (Improvement and Spatial Extension of the European Fish Index, EC 044096) and the EU FP7
470 Project MARS (Managing Aquatic ecosystems and water Resources under multiple Stress,
471 Contract No. 603378). Petr Zajicek received an ArcMap 10.2.2 license within the framework
472 of the ESRI educational program, ESRI Deutschland GmbH. We are thankful to two
473 anonymous reviewers for valuable comments and suggestions that helped to improve the
474 manuscript.

475 **References**

- 476 Akaike, H., 1981. Likelihood of a model and information criteria. *Journal of Econometrics* 16, 3–14.
477 doi:10.1016/0304-4076(81)90071-3
- 478 Aparicio, E., Carmona-Catot, G., Moyle, P.B., García-Berthou, E., 2011. Development and evaluation
479 of a fish-based index to assess biological integrity of Mediterranean streams. *Aquatic*
480 *Conserv. Mar. Freshw. Ecosyst.* 21, 324–337. doi:10.1002/aqc.1197
- 481 Barton, K., 2016. MuMIn: Multi-Model Inference [WWW Document]. R package version 1.15.6. URL
482 <https://CRAN.R-project.org/package=MuMIn> (accessed 6.25.16).
- 483 Bates, D., Mächler, M., Bolker, B., Walker, S., 2015. Fitting Linear Mixed-Effects Models using lme4.
484 *Journal of Statistical Software* 67. doi:10.18637/jss.v067.i01
- 485 Beier, U., Degerman, E., Melcher, A., Rogers, C., Wirlöf, H., 2007. Processes of collating a European
486 fisheries database to meet the objectives of the European Union Water Framework Directive.
487 *Fisheries Management and Ecology* 14, 407–416. doi:10.1111/j.1365-2400.2007.00579.x
- 488 Benitez, J.-P., Matondo, B.N., Dierckx, A., Ovidio, M., 2015. An overview of potamodromous fish
489 upstream movements in medium-sized rivers, by means of fish passes monitoring. *Aquat Ecol*
490 49, 481–497. doi:10.1007/s10452-015-9541-4
- 491 Berg, R., Gaumert, T., Kämmerleit, M., Klinger, H., Lemcke, R., Leuner, E., Wolter, C., Dußling, U.,
492 2004. The German river typology with respect to fish-faunistic reference conditions, in:
493 Steinberg, C., Calmano, W., Klapper, H., Wilken, R.-D. (Eds.), *Handbuch Angewandte*
494 *Limnologie*. Ecomed-Verlag, Landsberg, pp. 12–17.
- 495 Blondel, J., 2003. Guilds or functional groups: does it matter? *Oikos* 100, 223–231.
496 doi:10.1034/j.1600-0706.2003.12152.x

497 Bohlin, T., Hamrin, S., Heggberget, T., Rasmussen, G., Saltveit, S., 1989. Electrofishing — theory and
498 practice with special emphasis on salmonids. *Hydrobiologia* 173, 9–43.
499 doi:10.1007/BF00008596

500 Casselman, J., Lewis, C., 1996. Habitat requirements of Northern pike (*Esox lucius*). *Canadian Journal*
501 *of Fisheries and Aquatic Sciences* 53, 161–174.

502 Clark, S.J., Jackson, J.R., Lochmann, S.E., 2007. A Comparison of Shoreline Seines with Fyke Nets for
503 Sampling Littoral Fish Communities in Floodplain Lakes. *North American Journal of Fisheries*
504 *Management* 27, 676–680. doi:10.1577/M06-197.1

505 Council Directive 92/43/EEC, 1994. Council Directive 92/43/EEC of 21 May 1992 on the conservation
506 of natural habitats and of wild fauna and flora. *Official Journal of the European Communities*
507 L 206, 7–50.

508 De Leeuw, J.J., Buijse, A.D., Haidvogel, G., Lapinska, M., Noble, R., Repecka, R., Virbickas, T.,
509 Wiśniewolski, W., Wolter, C., 2007. Challenges in developing fish-based ecological
510 assessment methods for large floodplain rivers. *Fisheries Management and Ecology* 14, 483–
511 494. doi:10.1111/j.1365-2400.2007.00576.x

512 Dembkowski, D.J., Wuellner, M.R., Willis, D.W., 2012. Sampling Glacial Lake Littoral Fish Assemblages
513 with Four Gears. *North American Journal of Fisheries Management* 32, 1160–1166.
514 doi:10.1080/02755947.2012.728176

515 Dettmers, J.M., Gutreuter, S., Wahl, D.H., Soluk, D.A., 2001a. Patterns in abundance of fishes in main
516 channels of the upper Mississippi River system. *Can. J. Fish. Aquat. Sci.* 58, 933–942.
517 doi:10.1139/f01-046

518 Dettmers, J.M., Wahl, D.H., Soluk, D.A., Gutreuter, S., 2001b. Life in the fast lane: fish and foodweb
519 structure in the main channel of large rivers. *Journal of the North American Benthological*
520 *Society* 20, 255–265. doi:10.2307/1468320

521 Dußling, U., 2009. *Handbuch zu fiBS, Schriftenreihe des Verbandes Deutscher*
522 *Fischereiverwaltungsbeamter und Fischereiwissenschaftler e.V. Verband Deutscher*
523 *Fischereiverwaltungsbeamter und Fischereiwissenschaftler e.V., Offenbach am Main.*

524 Dußling, U., Bischoff, A., Haberbosch, R., Hoffmann, A., Klinger, H., Wolter, C., Wysujack, K., Berg, R.,
525 2004a. The fish-based assessment system - description of the German approach, in:
526 Steinberg, C., Calmano, W., Klapper, H., Wilken, R.-D. (Eds.), *Handbuch Angewandte*
527 *Limnologie. Ecomed-Verlag, Landsberg*, pp. 27–38.

528 Dußling, U., Bischoff, A., Haberbosch, R., Hoffmann, A., Klinger, H., Wolter, C., Wysujack, K., Berg, R.,
529 2004b. Ecological characters of river fish species occurring in Germany, in: Steinberg, C.,
530 Calmano, W., Klapper, H., Wilken, R.-D. (Eds.), *Handbuch Angewandte Limnologie. Ecomed-*
531 *Verlag, Landsberg*, pp. 4–12.

532 EFI + Consortium, 2009. Manual for the application of the new European Fish Index – EFI+. A fish-
533 based method to assess the ecological status of European running waters in support of the
534 Water Framework Directive.

535 Eggleton, M.A., Jackson, J.R., Lubinski, B.J., 2010. Comparison of gears for sampling littoral-zone
536 fishes in floodplain lakes of the lower White River, Arkansas. *North Amer. J. of Fisheries*
537 *Mgmt* 30, 928–939. doi:10.1577/M09-127.1

538 Erős, T., Tóth, B., Sevcsik, A., Schmera, D., 2008. Comparison of Fish Assemblage Diversity in Natural
539 and Artificial Rip-Rap Habitats in the Littoral Zone of a Large River (River Danube, Hungary).
540 *International Review of Hydrobiology* 93, 88–105. doi:10.1002/iroh.200710976

541 Fladung, E., 2002. Untersuchungen zum adulten Fischbestand im Hauptstrom (Fahrrinne) der
542 Mittelalbe. *Zeitschrift für Fischkunde, Supplement* 1, 121–131.

543 Flotemersch, J.E., Stribling, J.B., Hughes, R.M., Reynolds, L., Paul, M.J., Wolter, C., 2011. Site length
544 for biological assessment of boatable rivers. *River Res. Applic.* 27, 520–535.
545 doi:10.1002/rra.1367

546 Fullerton, A.H., Burnett, K.M., Steel, E.A., Flitcroft, R.L., Pess, G.R., Feist, B.E., Torgersen, C.E., Miller,
547 D.J., Sanderson, B. I., 2010. Hydrological connectivity for riverine fish: measurement

548 challenges and research opportunities. *Freshwater Biology* 55, 2215–2237.
 549 doi:10.1111/j.1365-2427.2010.02448.x
 550 Galat, D.L., Zweimüller, I., 2001. Conserving large-river fishes: is the highway analogy an appropriate
 551 paradigm? *Journal of the North American Benthological Society* 20, 266–279.
 552 doi:10.2307/1468321
 553 Goffaux, D., Grenouillet, G., Kestemont, P., 2005. Electrofishing versus gillnet sampling for the
 554 assessment of fish assemblages in large rivers. *Archiv für Hydrobiologie* 162, 73–90.
 555 doi:10.1127/0003-9136/2005/0162-0073
 556 Gonzales, R., Griffin, D., 2004. Measuring individuals in a social environment: Conceptualizing dyadic
 557 and group interaction, in: Sansone, C., Morf, C.C., Panter, A.T. (Eds.), *The Sage Handbook of*
 558 *Methods in Social Psychology*. Sage Publications, Inc., California, London, New Delhi, pp.
 559 313–334.
 560 Gutreuter, S., Burkhardt, R., Lubinski, K., 1995. Long Term Resource Monitoring Program Procedures:
 561 Fish Monitoring (No. LTRMP 95-P002-1). National Biological Service, Environmental
 562 Management Technical Center, Onalaska, Wisconsin.
 563 Heino, J., Schmera, D., Erős, T., 2013. A macroecological perspective of trait patterns in stream
 564 communities. *Freshw Biol* 58, 1539–1555. doi:10.1111/fwb.12164
 565 Hothorn, T., Bretz, F., Westfall, P., Heiberger, R.M., Schuetzenmeister, A., Scheibe, S., 2016.
 566 multcomp: Simultaneous Inference in General Parametric Models [WWW Document]. R
 567 package version 1.4-5. URL <https://cran.r-project.org/web/packages/multcomp/index.html>
 568 (accessed 6.23.16).
 569 Illies, J., 1961. Versuch einer allgemeinen biozönotischen Gliederung der Fließgewässer. *Int. Revue*
 570 *ges. Hydrobiol. Hydrogr.* 46, 205–213. doi:10.1002/iroh.19610460205
 571 Jacobsen, L., Perrow, M.R., 1998. Predation risk from piscivorous fish influencing the diel use of
 572 macrophytes by planktivorous fish in experimental ponds. *Ecol Freshwater Fish* 7, 78–86.
 573 doi:10.1111/j.1600-0633.1998.tb00174.x
 574 Kail, J., Wolter, C., 2011. Analysis and evaluation of large-scale river restoration planning in Germany
 575 to better link river research and management. *River Res. Applic.* 27, 985–999.
 576 doi:10.1002/rra.1382
 577 Kemp, P.S., O’hanley, J.R., 2010. Procedures for evaluating and prioritising the removal of fish
 578 passage barriers: a synthesis. *Fisheries Management and Ecology* 17, 297–322.
 579 doi:10.1111/j.1365-2400.2010.00751.x
 580 Kennedy, G., Strange, C., 1981. Efficiency of Electric Fishing for Salmonids in Relation to River Width.
 581 *Fisheries Management* 12, 55–60.
 582 Kucera-Hirzinger, V., Schludermann, E., Zornig, H., Weissenbacher, A., Schabuss, M., Schiemer, F.,
 583 2008. Potential effects of navigation-induced wave wash on the early life history stages of
 584 riverine fish. *Aquat. Sci.* 71, 94–102. doi:10.1007/s00027-008-8110-5
 585 Kuznetsova, A., Brockhoff, B., Christensen, H.B., 2016. lmerTest: Tests in Linear Mixed Effects Models
 586 [WWW Document]. R package version 2.0-32. URL [https://CRAN.R-](https://CRAN.R-project.org/package=lmerTest)
 587 [project.org/package=lmerTest](https://CRAN.R-project.org/package=lmerTest) (accessed 6.23.16).
 588 Lechner, A., Keckeis, H., Humphries, P., 2016. Patterns and processes in the drift of early
 589 developmental stages of fish in rivers: a review. *Rev Fish Biol Fisheries* 26, 471–489.
 590 doi:10.1007/s11160-016-9437-y
 591 Lechner, A., Keckeis, H., Schludermann, E., Loisl, F., Humphries, P., Glas, M., Tritthart, M., Habersack,
 592 H., 2014. Shoreline configurations affect dispersal patterns of fish larvae in a large river. *ICES*
 593 *J. Mar. Sci.* 71, 930–942. doi:10.1093/icesjms/fst139
 594 Lewin, W.-C., Mehner, T., Ritterbusch, D., Brämick, U., 2014. The influence of anthropogenic
 595 shoreline changes on the littoral abundance of fish species in German lowland lakes varying
 596 in depth as determined by boosted regression trees. *Hydrobiologia* 724, 293–306.
 597 doi:10.1007/s10750-013-1746-8
 598 Lintermans, M., 2016. Finding the needle in the haystack: comparing sampling methods for detecting
 599 an endangered freshwater fish. *Mar. Freshwater Res.* 67, 1740–1749. doi:10.1071/MF14346

600 Loisl, F., Singer, G., Keckeis, H., 2013. Method-integrated fish assemblage structure at two spatial
601 scales along a free-flowing stretch of the Austrian Danube. *Hydrobiologia* 729, 77–94.
602 doi:10.1007/s10750-013-1588-4

603 Lynch, W.E.J., Johnson, D.L., 1989. Influences of Interstice Size, Shade, and Predators on the Use of
604 Artificial Structures by Bluegills. *North American Journal of Fisheries Management* 9, 219–
605 225. doi:10.1577/1548-8675(1989)009<0219:IOISSA>2.3.CO;2

606 Lyon, J.P., Bird, T., Nicol, S., Kearns, J., O’Mahony, J., Todd, C.R., Cowx, I.G., Bradshaw, C.J.A., 2014.
607 Efficiency of electrofishing in turbid lowland rivers: implications for measuring temporal
608 change in fish populations. *Can. J. Fish. Aquat. Sci.* 71, 878–886. doi:10.1139/cjfas-2013-0287

609 Magurran, A.E., Henderson, P.A., 2003. Explaining the excess of rare species in natural species
610 abundance distributions. *Nature* 422, 714–716. doi:10.1038/nature01547

611 Morales, M., R Development Core Team, Murdoch, D., 2012. *sciplot: Scientific Graphing Functions for*
612 *Factorial Designs* [WWW Document]. R package version 1.1-0. URL [https://CRAN.R-](https://CRAN.R-project.org/package=sciplot)
613 [project.org/package=sciplot](https://CRAN.R-project.org/package=sciplot) (accessed 10.15.15).

614 Morandi, B., Piégay, H., Lamouroux, N., Vaudor, L., 2014. How is success or failure in river restoration
615 projects evaluated? Feedback from French restoration projects. *Journal of Environmental*
616 *Management* 137, 178–188. doi:10.1016/j.jenvman.2014.02.010

617 Morris, E.K., Caruso, T., Buscot, F., Fischer, M., Hancock, C., Maier, T.S., Meiners, T., Müller, C.,
618 Obermaier, E., Prati, D., Socher, S.A., Sonnemann, I., Wäschke, N., Wubet, T., Wurst, S., Rillig,
619 M.C., 2014. Choosing and using diversity indices: insights for ecological applications from the
620 German Biodiversity Exploratories. *Ecol Evol* 4, 3514–3524. doi:10.1002/ece3.1155

621 Nakagawa, S., Schielzeth, H., 2013. A general and simple method for obtaining R² from generalized
622 linear mixed-effects models. *Methods Ecol Evol* 4, 133–142. doi:10.1111/j.2041-
623 210x.2012.00261.x

624 Neebling, T.E., Quist, M.C., 2011. Comparison of Boat Electrofishing, Trawling, and Seining for
625 Sampling Fish Assemblages in Iowa’s Nonwadeable Rivers. *North American Journal of*
626 *Fisheries Management* 31, 390–402. doi:10.1080/02755947.2011.576198

627 Noble, R. a. A., Cowx, I.G., Goffaux, D., Kestemont, P., 2007. Assessing the health of European rivers
628 using functional ecological guilds of fish communities: standardising species classification and
629 approaches to metric selection. *Fisheries Management and Ecology* 14, 381–392.
630 doi:10.1111/j.1365-2400.2007.00575.x

631 Penczak, T., Jakubowski, H., 1990. Drawbacks of electric fishing in rivers, in: Cowx, I.G. (Ed.),
632 *Developments in Electric Fishing*. Blackwell, Oxford, pp. 115–122.

633 Piper, A.T., Manes, C., Siniscalchi, F., Marion, A., Wright, R.M., Kemp, P.S., 2015. Response of
634 seaward-migrating European eel (*Anguilla anguilla*) to manipulated flow fields. *Proc. R. Soc. B*
635 282, 20151098. doi:10.1098/rspb.2015.1098

636 Poesch, M.S., 2014. Developing Standardized Methods for Sampling Freshwater Fishes with Multiple
637 Gears: Effects of Sampling Order versus Sampling Method. *Transactions of the American*
638 *Fisheries Society* 143, 353–362. doi:10.1080/00028487.2013.860047

639 Poikane, S., Zampoukas, N., Borja, A., Davies, S.P., van de Bund, W., Birk, S., 2014. Intercalibration of
640 aquatic ecological assessment methods in the European Union: Lessons learned and way
641 forward. *Environmental Science & Policy* 44, 237–246. doi:10.1016/j.envsci.2014.08.006

642 Poos, M.S., Mandrak, N.E., McLaughlin, R.L., 2007. The effectiveness of two common sampling
643 methods for assessing imperilled freshwater fishes. *Journal of Fish Biology* 70, 691–708.
644 doi:10.1111/j.1095-8649.2007.01349.x

645 Porreca, A.P., Pederson, C.L., Laursen, J.R., Colombo, R.E., 2013. A comparison of electrofishing
646 methods and fyke netting to produce reliable abundance and size metrics. *Journal of*
647 *Freshwater Ecology* 28, 585–590. doi:10.1080/02705060.2013.810555

648 R Development Core Team, 2016. *R: A Language and Environment for Statistical Computing* [WWW
649 Document]. R Foundation For Statistical Computing, Vienna, Austria. URL [https://www.r-](https://www.r-project.org/)
650 [project.org/](https://www.r-project.org/) (accessed 6.21.16).

651 Radinger, J., Wolter, C., 2014. Patterns and predictors of fish dispersal in rivers. *Fish Fish* 15, 456–
652 473. doi:10.1111/faf.12028

653 Randall, R.G., Minns, C.K., Cairns, V.W., Moore, J.E., 1996. The relationship between an index of fish
654 production and submerged macrophytes and other habitat features at three littoral areas in
655 the Great Lakes. *Can. J. Fish. Aquat. Sci.* 53, 35–44. doi:10.1139/f95-271

656 Scharf, J., Brämick, U., Fredrich, F., Rothe, U., Schuhr, H., Tautenhahn, M., Wolter, C., Zahn, S., 2011.
657 Fische in Brandenburg - Aktuelle Kartierung und Beschreibung der märkischen Fischfauna.

658 Somerfield, P.J., Clarke, K.R., Warwick, R.M., 2008. Simpson Index, in: Fath, B.D. (Ed.), *Encyclopedia of*
659 *Ecology*. Academic Press, Oxford, pp. 3252–3255.

660 Spellerberg, I.F., 2008. Shannon–Wiener Index, in: Fath, B.D. (Ed.), *Encyclopedia of Ecology*. Academic
661 Press, Oxford, pp. 3249–3252.

662 Spellerberg, I.F., Fedor, P.J., 2003. A tribute to Claude Shannon (1916–2001) and a plea for more
663 rigorous use of species richness, species diversity and the ‘Shannon–Wiener’ Index. *Global*
664 *Ecology and Biogeography* 12, 177–179. doi:10.1046/j.1466-822X.2003.00015.x

665 Strayer, D.L., Findlay, S.E.G., 2010. Ecology of freshwater shore zones. *Aquatic Sciences* 72, 127–163.
666 doi:10.1007/s00027-010-0128-9

667 Szalóky, Z., György, Á.I., Tóth, B., Sevcsik, A., Specziár, A., Csányi, B., Szekeres, J., Erős, T., 2014.
668 Application of an electrified benthic frame trawl for sampling fish in a very large European
669 river (the Danube River) – Is offshore monitoring necessary? *Fisheries Research* 151, 12–19.
670 doi:10.1016/j.fishres.2013.12.004

671 Tokeshi, M., 1993. Species Abundance Patterns and Community Structure. *Advances in Ecological*
672 *Research* 24, 111–186. doi:10.1016/S0065-2504(08)60042-2

673 Tukey, J.W., 1949. Comparing individual means in the analysis of variance. *Biometrics* 5, 99–114.

674 Ward, J.V., Tockner, K., Arscott, D.B., Claret, C., 2002. Riverine landscape diversity. *Freshwater*
675 *Biology* 47, 517–539. doi:10.1046/j.1365-2427.2002.00893.x

676 Weaver, M.J., Magnuson, J.J., Clayton, M.K., 1997. Distribution of littoral fishes in structurally
677 complex macrophytes. *Can. J. Fish. Aquat. Sci.* 54, 2277–2289. doi:10.1139/cjfas-54-10-2277

678 Welcomme, R.L., Winemiller, K.O., Cowx, I.G., 2006. Fish environmental guilds as a tool for
679 assessment of ecological condition of rivers. *River Res. Applic.* 22, 377–396.
680 doi:10.1002/rra.914

681 Wiley, M.L., Tsai, C.-F., 1983. The Relative Efficiencies of Electrofishing vs. Seines in Piedmont
682 Streams of Maryland. *North American Journal of Fisheries Management* 3, 243–253.
683 doi:10.1577/1548-8659(1983)3<243:TREOEV>2.0.CO;2

684 Wolter, C., Bischoff, A., 2001. Seasonal changes of fish diversity in the main channel of the large
685 lowland River Oder. *Regul. Rivers: Res. Mgmt.* 17, 595–608. doi:10.1002/rrr.645

686 Wolter, C., Bischoff, A., Faller, M., Schomaker, C., Wysujack, K., 2004. Sampling design and site
687 selection in large rivers, in: Steinberg, C., Calmano, W., Klapper, H., Wilken, R.-D. (Eds.),
688 *Handbuch Angewandte Limnologie*. Ecomed-Verlag, Landsberg, pp. 38–57.

689 Wolter, C., Freyhof, J., 2004. Diel distribution patterns of fishes in a temperate large lowland river.
690 *Journal of Fish Biology* 64, 632–642. doi:10.1111/j.1095-8649.2004.00327.x

691 Wolter, C., Lorenz, S., Scheunig, S., Lehmann, N., Schomaker, C., Nastase, A., García de Jalón, D.,
692 Marzin, A., Lorenz, A.W., Krakova, M., Brabec, K., Noble, R., 2013. Review on ecological
693 response to hydromorphological degradation and restoration (No. D1.3).

694 Zitek, A., Schmutz, S., Unfer, G., Ploner, A., 2004. Fish drift in a Danube sidearm-system: I. Site-, inter-
695 and intraspecific patterns. *Journal of Fish Biology* 65, 1319–1338. doi:10.1111/j.0022-
696 1112.2004.00533.x

697 Zuur, A.F., Ieno, E.N., 2016. A protocol for conducting and presenting results of regression-type
698 analyses. *Methods Ecol Evol* 7, 636–645. doi:10.1111/2041-210X.12577

699 Zuur, A.F., Ieno, E.N., Walker, N.J., Saveliev, A.A., Smith, G.M., 2009. *Mixed effects models and*
700 *extensions in ecology with R*. Springer, New York NY.

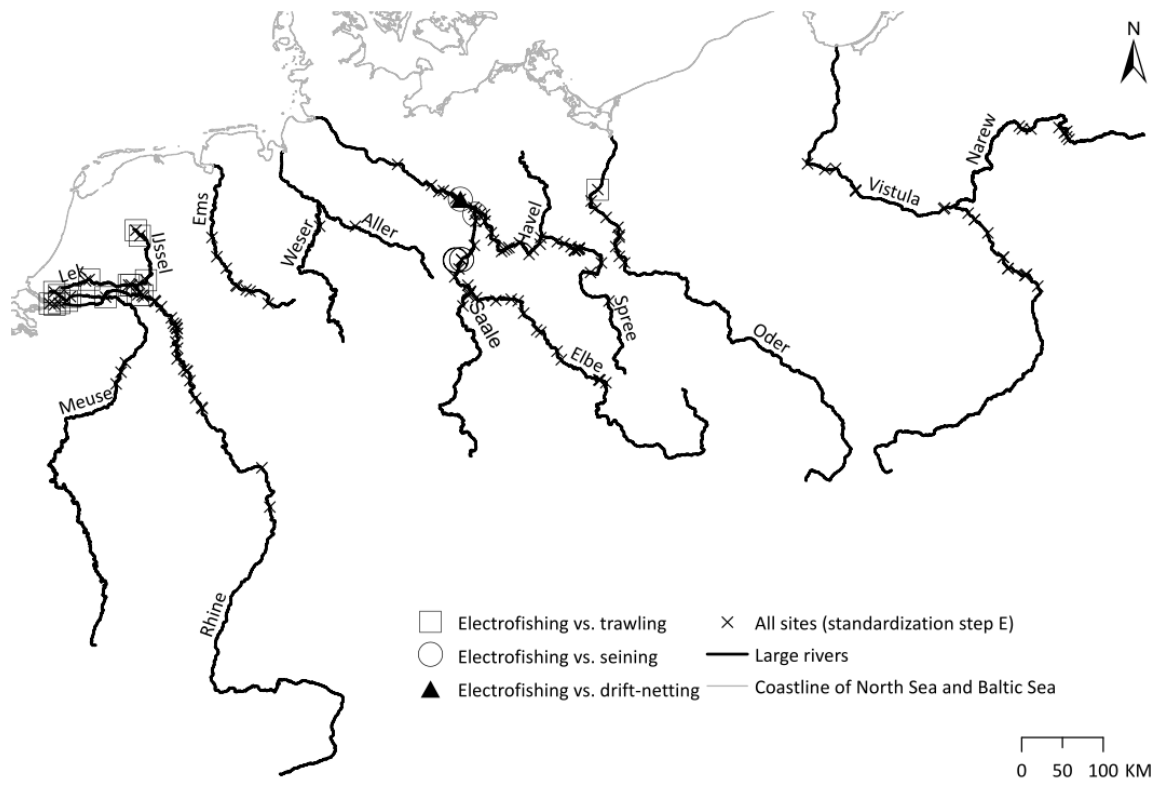
702
703
704
705
706
707
708

Tables

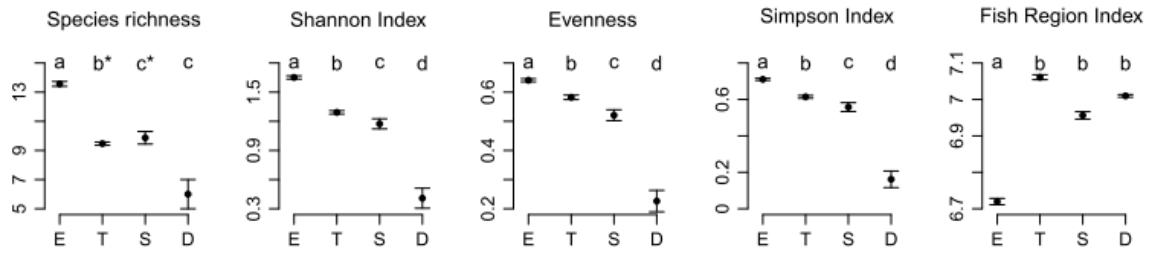
Table 1. Species numbers and ratios of fishes captured with each gear (E = electrofishing; T = trawling; S = seining; D = drift-netting) for the preliminary comparison of all gears and for standardized comparisons of electrofishing versus each additional gear. Sam, Sp, Excl and Fi = total numbers of samplings, of species, of exclusive species and of captured fish (=total catch), respectively. EURY, RH, LITH, PHYT, PSAM and POT = eurytopic, rheophilic, lithophilic, phytophilic, psammophilic and potamal guilds, respectively. MIG = migratory species and HD = species listed in annexes of the Habitat Directive. “n” refers to the number of species and “PROP” refers to the ratio of fishes in the total catch captured with the respective gear

Gear	Sam	Sp	Excl	Fi	EURY		RH		LITH		PHYT		PSAM		POT		MIG		HD	
					[n]	[PROP]	[n]	[PROP]	[n]	[PROP]	[n]	[PROP]	[n]	[PROP]	[n]	[PROP]	[n]	[PROP]	[n]	[PROP]
Preliminary comparison of all gears																				
E	512	62	22	304155	20	71.8	32	27.5	19	9.7	13	6	4	5.7	6	6.9	15	15.7	14	2.8
T	297	40	3	177924	16	90.3	21	9.7	13	0.2	8	14.5	2	2.1	5	64.2	16	8.3	9	0.3
S	38	26	1	21219	11	90.3	11	9.3	5	2.5	8	21.5	2	3.3	4	74.0	6	1.9	6	1.9
D	2	8	0	295	6	99.3	2	0.7	0	0	1	90.8	0	0	4	93.2	1	0.3	0	0
Standardized gear comparisons																				
E	162	41	7	74393	17	69.5	17	30.1	7	2.8	11	6.5	3	1.5	5	5.1	12	20.1	7	3.1
T	284	40	6	174647	16	90.7	21	9.3	13	0.2	8	14.5	2	2	5	64.3	16	8.1	9	0.3
E	56	30	13	30238	13	66.7	15	33.1	7	7.5	9	8.3	4	5.6	5	15.1	7	11.1	5	2.3
S	22	20	1	9151	10	93.8	9	6.2	4	2.3	4	28	2	0.1	4	71.4	5	2.3	2	1.9
E	8	17	10	3897	9	66.1	8	33.9	4	6.1	2	3	2	5.4	4	7.3	5	10.9	3	3.6
D	2	10	1	295	6	99.3	2	0.7	0	0	1	90.8	0	0	4	93.2	1	0.3	0	0

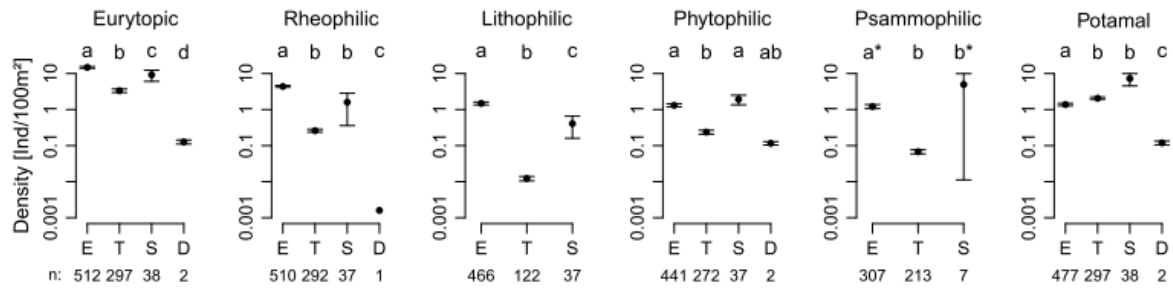
709



711 **Figure 1.** Location of sampling sites
712
713

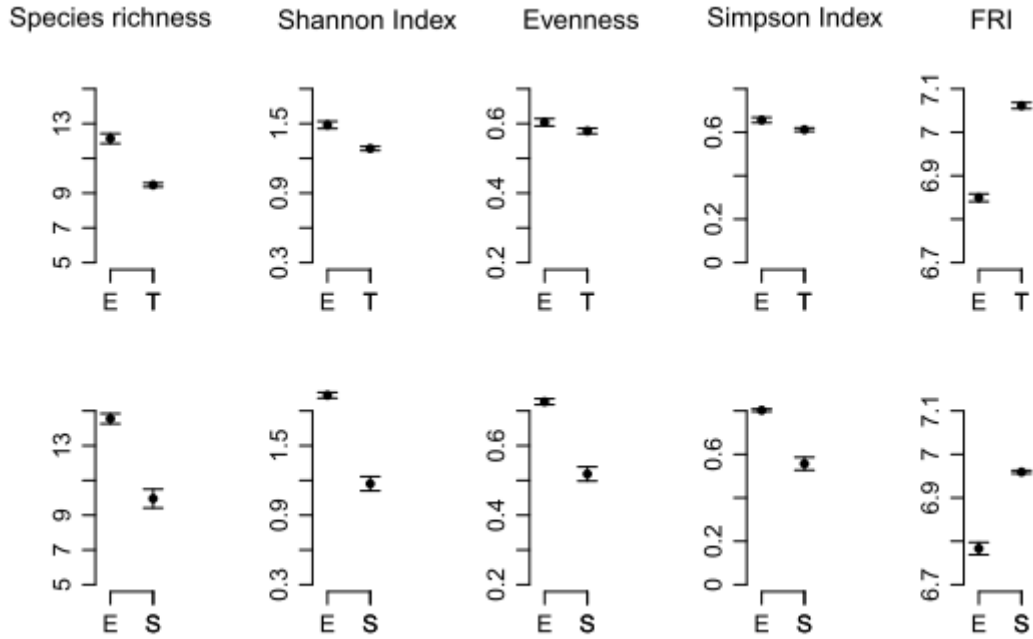


714
715 **Figure 2.** Biodiversity as estimated across 849 samples in European large rivers (E =
716 electrofishing [512 samples]; T = trawling [297], S = seining [38], D = drift-netting [2]).
717 Different lower case letters indicate significant differences; *note that species richness
718 estimated by T is significantly higher compared to S when accounting for unequal sample
719 sizes and random effects in a mixed effects model. D has a little sample size which requires
720 cautious interpretation. Y-axis is log-scaled, mean and +/- standard errors (Table S6) are
721 shown
722

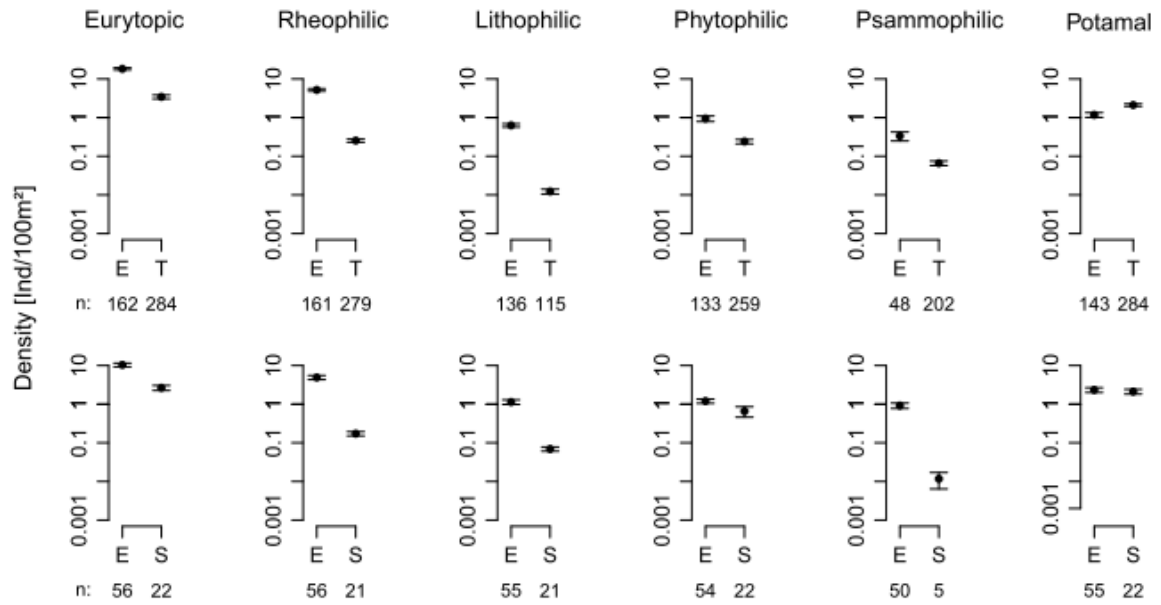


723
724
725
726
727
728
729
730
731
732
733
734
735

Figure 3. Densities of selected guilds as estimated across 849 samples in European large rivers (E = electrofishing [512 samples]; T = trawling [297], S = seining [38], D = drift-netting [2]; sample sizes (n) differ between guilds and same gears due to non-catches of fish in some samplings). Different lower case letters indicate significant differences. *Note that the high average value for the psammophilic density determined with S is biased due to one outlier and log transformed density estimated with electrofishing is significantly higher as estimated with seining for the psammophilic guild when also accounting for unequal sample sizes and random effects in a mixed effects model. Y-axis is log-scaled, mean and +/- standard errors (Table S6) are shown. D has a little sample size which requires cautious interpretation and no species belonging to lithophilic and psammophilic guilds were caught with D

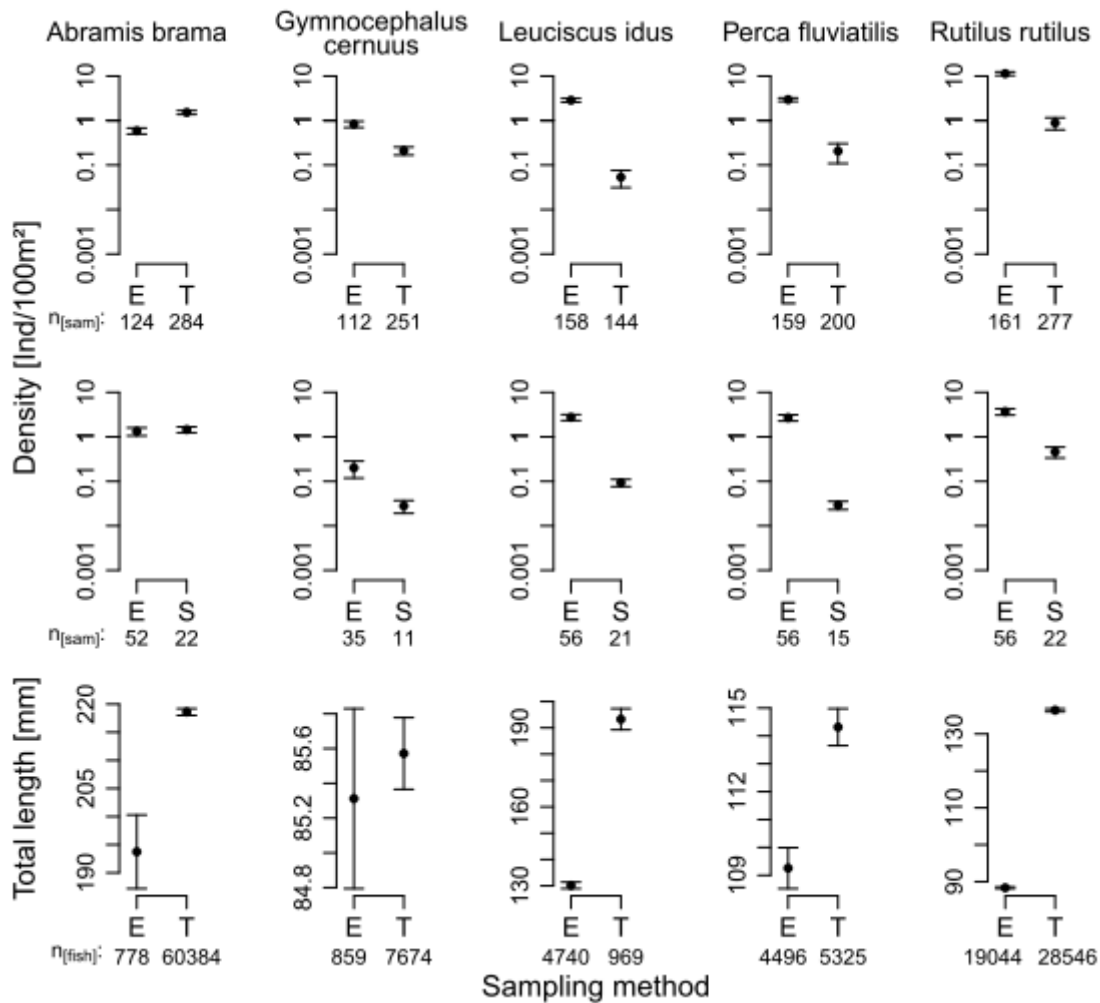


736
 737 **Figure 4.** Biodiversity indices as estimated in the standardized gear comparisons of
 738 electrofishing (E) vs. trawling (T) [samples: E = 162; T = 284] and E vs. seining (S) [E = 56;
 739 S = 22]. “FRI” = Fish Region Index. All differences are significant. Mean +/- standard errors
 740 (Tables S9, S11) are shown
 741



742
743
744
745
746
747
748
749

Figure 5. Densities of selected guilds as estimated in the standardized gear comparisons of electrofishing (E) vs. trawling (T) [samples : E = 162; T = 284] and E vs. seining (S) [E = 56; S = 22]. All differences are significant except the density of the potamal guild within the E vs. S comparison. Sample sizes (n) differ between guilds and same gears due to non-catches of fish belonging to the respective guild in some samplings. Y-axis is log-scaled, mean and +/- standard errors (Tables S9, S11) are shown



751

752

753

754

755

756

757

758

Figure 6. Densities and total lengths of common species as estimated in the standardized gear comparisons of electrofishing (E) vs. trawling (T) [samples: E = 162; T = 284] and densities of common species as estimated in the comparison of E vs. seining (S) [E = 56; S = 22]. Sample sizes ($n_{[sam]}$ = number of samplings) differ between species and gears due to non-catches of species in some samplings; $n_{[fish]}$ = number of measured fish. All differences are significant despite total lengths of *Gymnocephalus cernuus*. Y-axis is log-scaled concerning density-plots. Mean and +/- standard errors (Tables S14, S16, S18) are shown in all plots